



# ***Roles of nitrifiers in the removal of micropollutants during wastewater treatment processes***

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ISTC sustainability seminar

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# Acknowledgements



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Kathrin Fenner



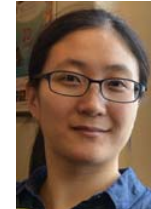
David Johnson



Damian Helbling



Michael Wagner



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Li-Jun Zhou

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Stefan Achermann



Rebekka Gulde



Andreas Maccagnan



Microbe - Environment Nexus



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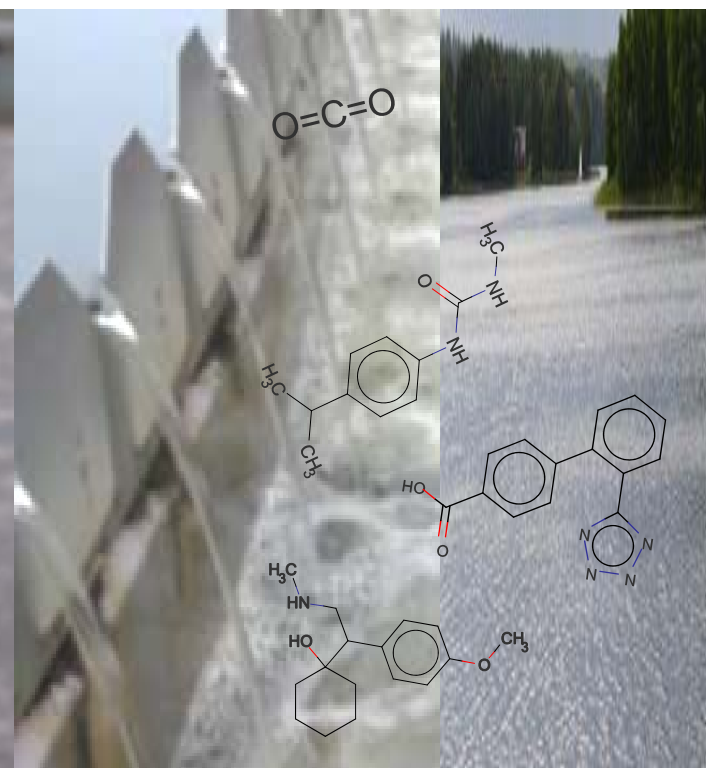
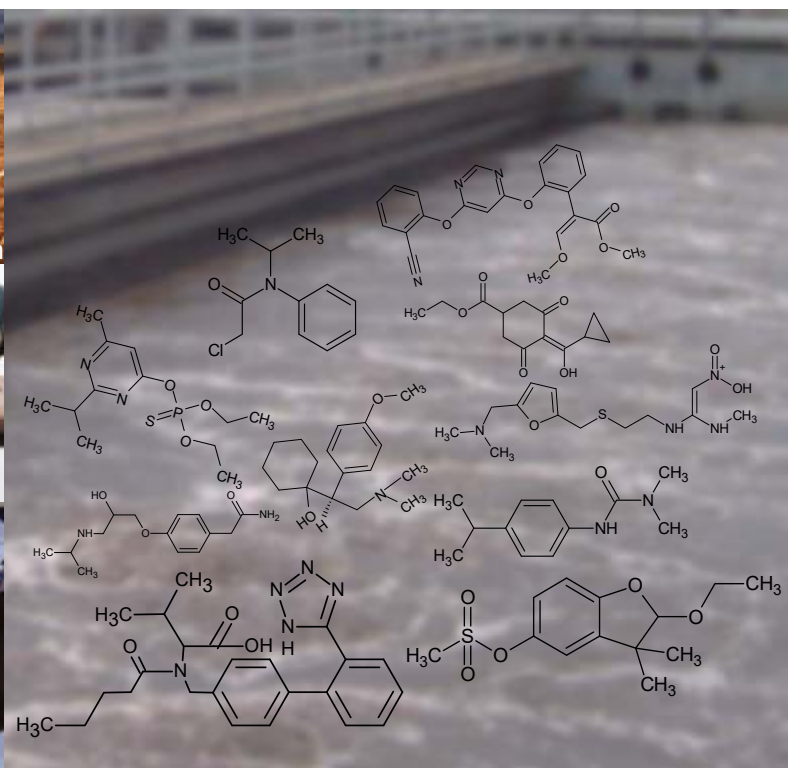
# Background

## Fate of micropollutants (MPs) in Wastewater Treatment Plants

MPs

Activated sludge communities

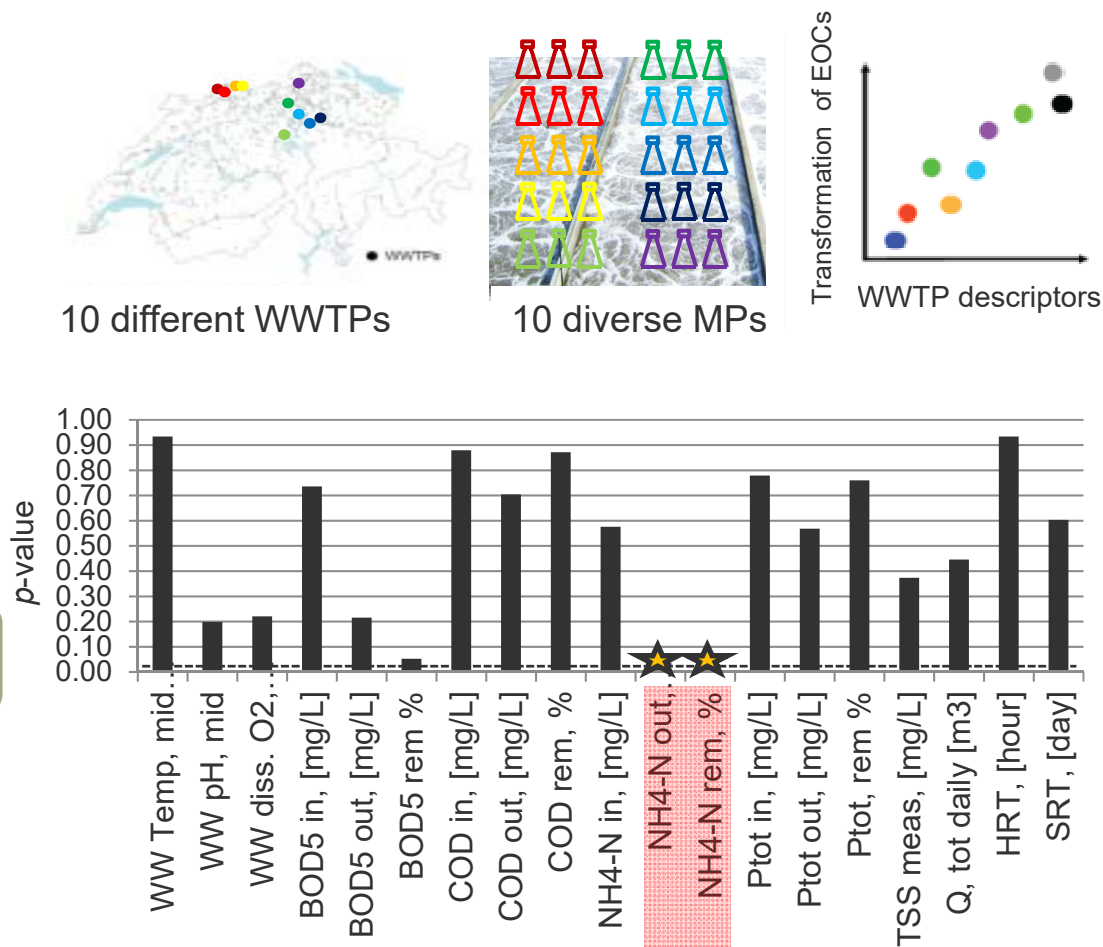
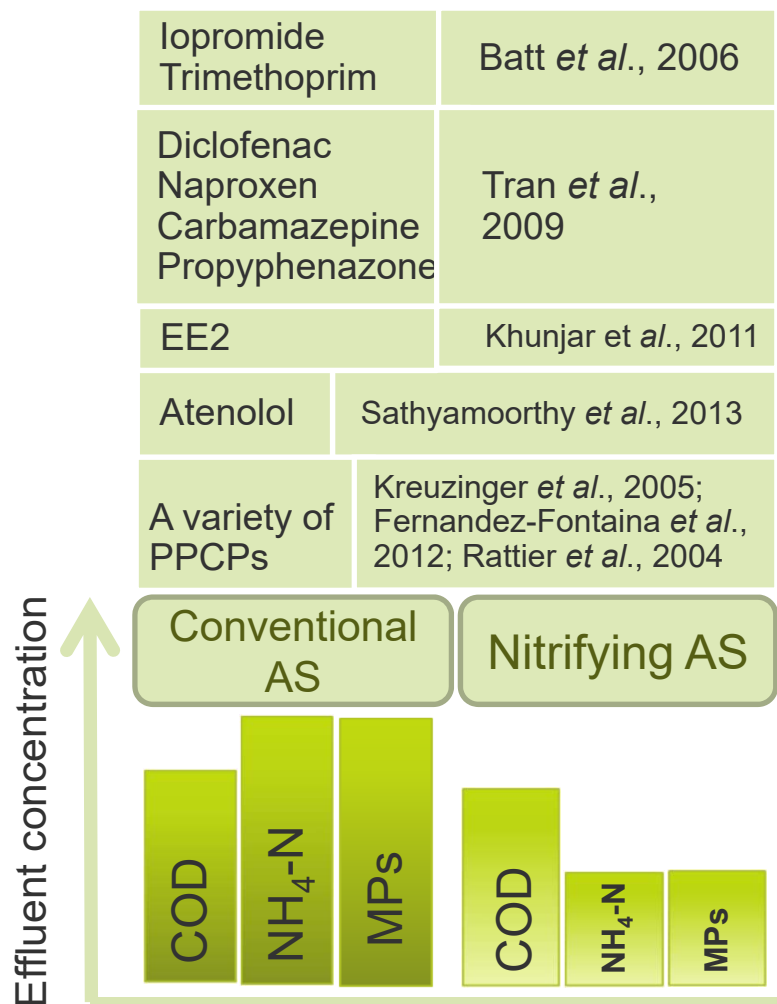
Discharge



- ? Persistent or biodegradable?
- ? Complete or incomplete transformation?
- ? Ecotoxicity of parent compounds and transformation products?

# Background

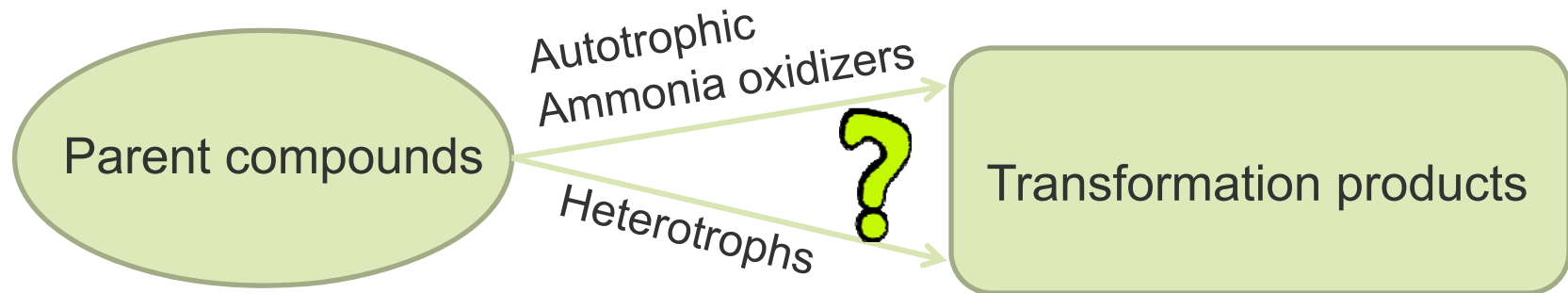
## Micropollutant biotransformation by activated sludge communities



(Helbling *et al.*, 2012. ES&T)

# Hypothesis

Causal relationship between ammonia oxidizers and MP biotransformation



## To test the hypothesis

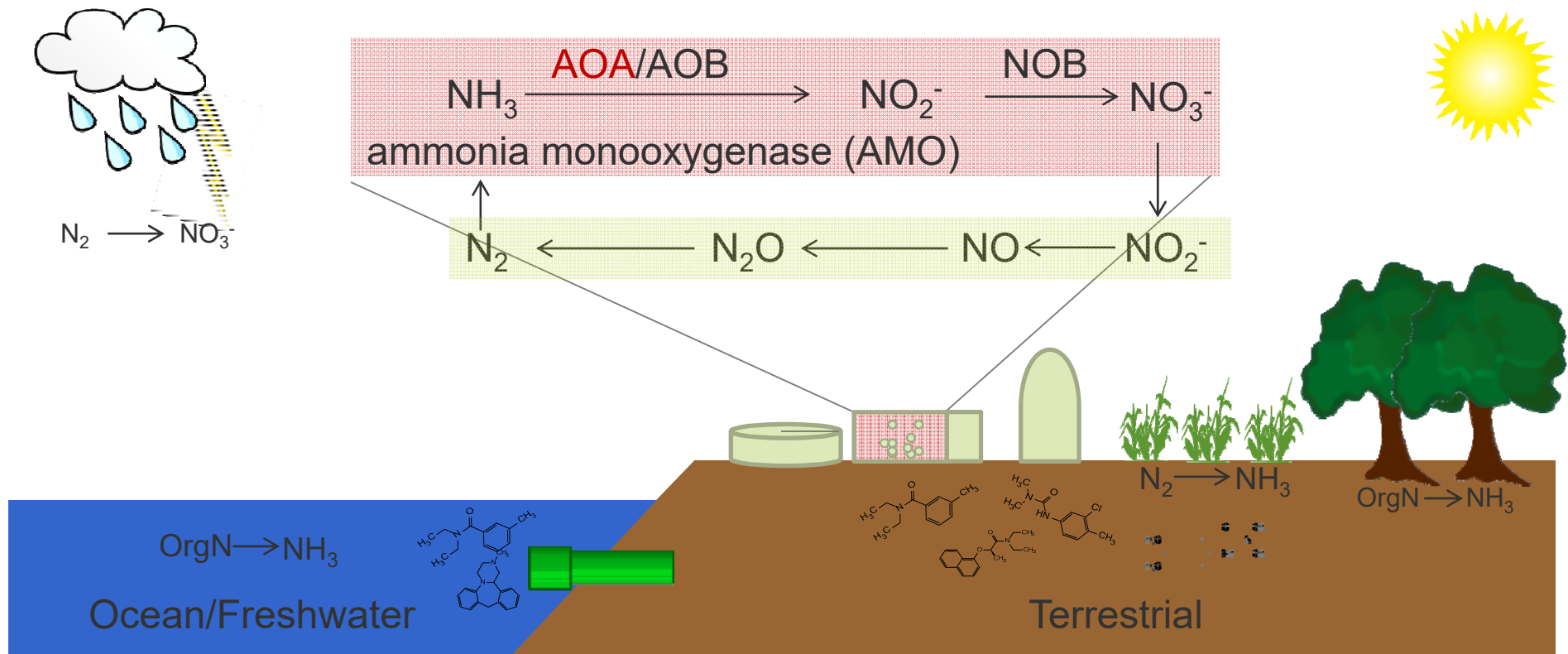
- ☐ Pure culture study
  - ✓ Ammonia-Oxidizing Bacteria (AOB)
  - ? Ammonia-Oxidizing Archaea (AOA)
- ☐ Inhibition study
- ☐ Removal in full-scale WWTPs



# MP biotransformation by an AOA pure culture

## Why AOA?

- Distribution and abundance
- Distinct physiological characteristics
- Unknown ecological significance in MP biotransformation



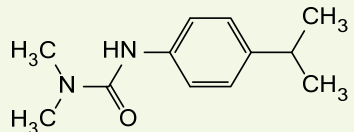
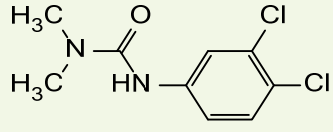
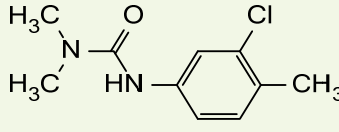
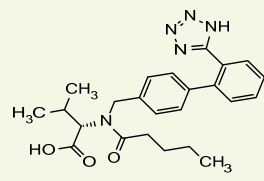
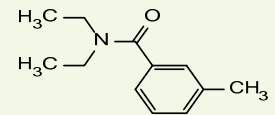
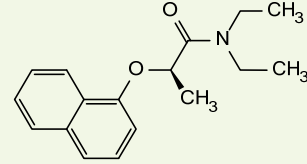
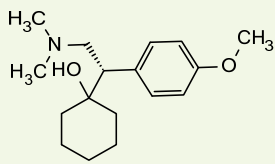
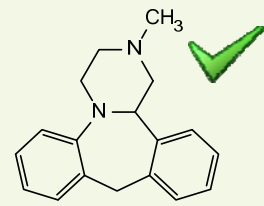
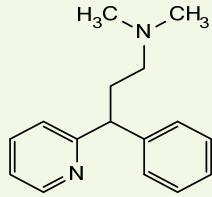
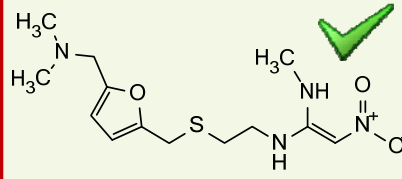
# MP biotransformation capability of AOA

- Tested ammonia oxidizers:

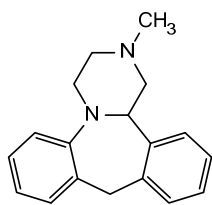
AOA: *Nitrososphaera gargensis*

AOB: *Nitrosomonas nitrosa* strain Nm90, Nm95

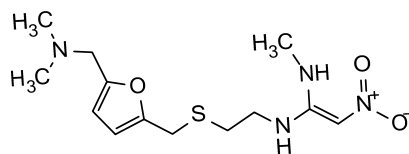
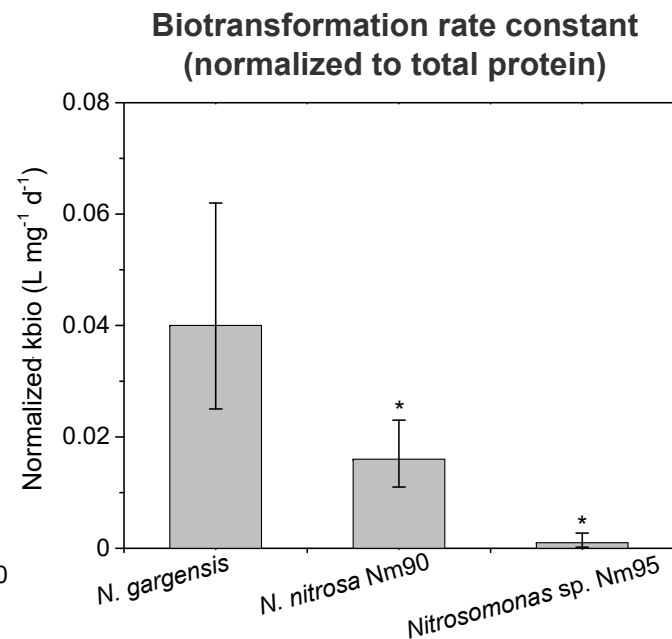
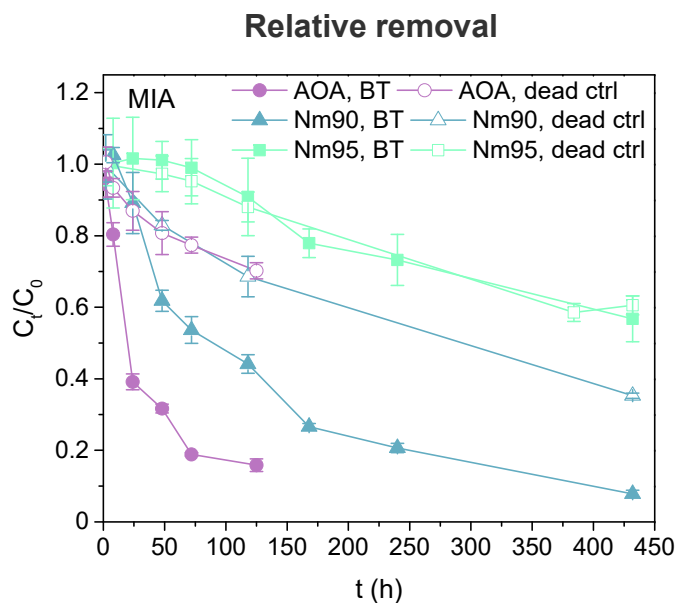
- Tested MPs (40 µg/L each ):

Phenyl ureas			
			
Isoproturon (ISO)	Diuron (DIU)	Chlortoluron (CHL)	
Tertiary amides			
			
Valsartan (VAL)	Diethyltoluamide (DEET)	Napropamide (NAP)	
Tertiary amines			
			
Venlafaxine (VEN)	Mianserin (MIA)	Pheniramine (PHE)	Ranitidine (RAN)

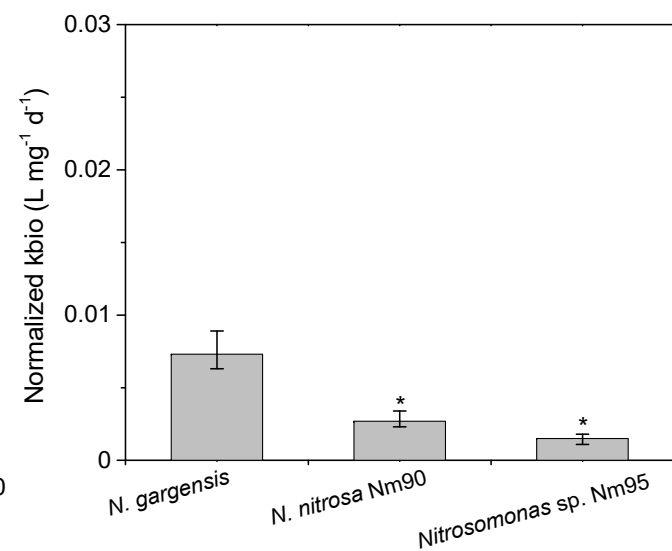
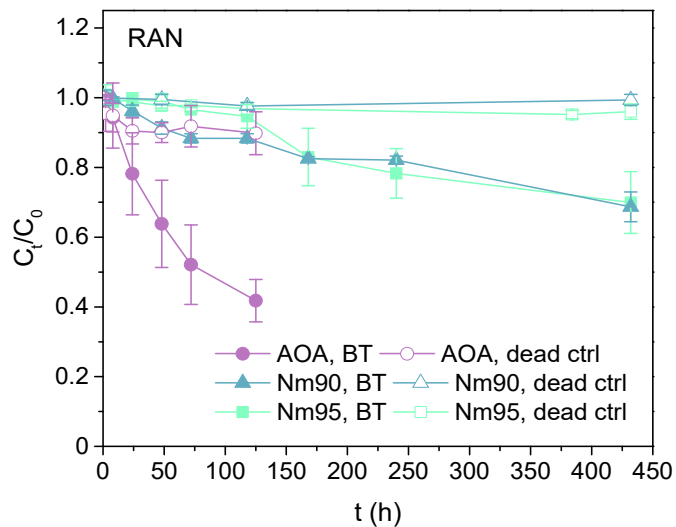
# Two MPs (MIA and RAN) were biotransformed by AOA



MIA



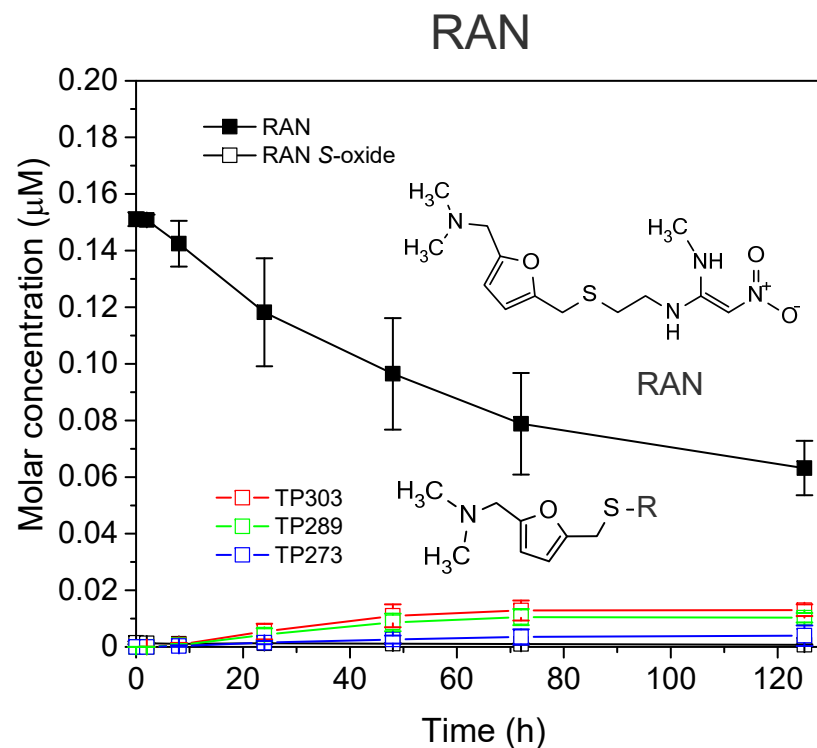
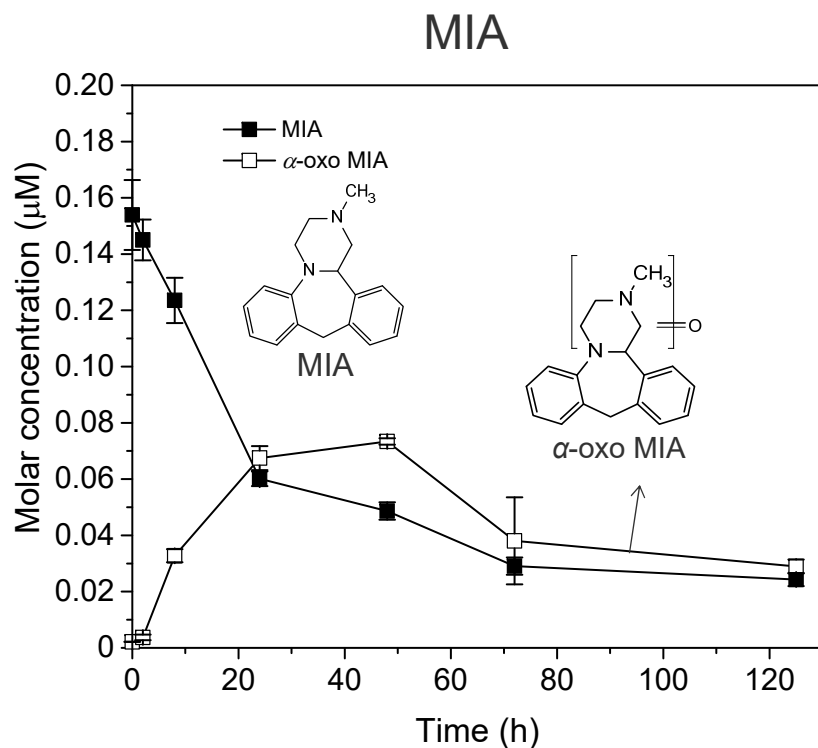
RAN





# Transformation product (TP) identification

- One TP ( $m/z$  279.1492) of MIA was determined to be  **$\alpha$ -oxo mianserin**.
- No *N*-oxidation (mianserin *N*-oxide) or *N*-demethylation (normianserin)
- No suspected RAN TP was detected, except RAN *S*-oxide
- Three RAN TPs from non target analysis



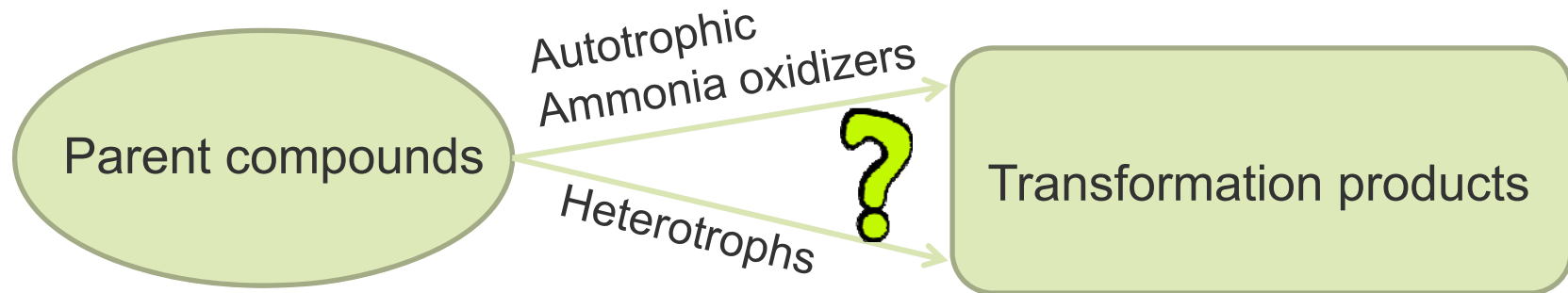
(TPs were semi-quantified according to peak areas)

## Summary — Pure culture study

- ✓ *N. gargensis* (an **AOA** strain) can biotransform two out of the ten tested MPs: **mianserin** (MIA) and **ranitidine** (RAN).
- ✓  **$\alpha$ -oxo mianserin** was the major MIA TP.
- ✓ Three TPs were identified for RAN from non-target analysis.
- ✓ Biotransformation of MIA and RAN by AOA was **cometabolic** .

# Hypothesis

Causal relationship between ammonia oxidizers and MP biotransformation



## To test the hypothesis

- ☐ Pure culture study
  - ✓ AOB
  - ✓ AOA
- ☐ Inhibition study
- ☐ Removal in full-scale WWTPs

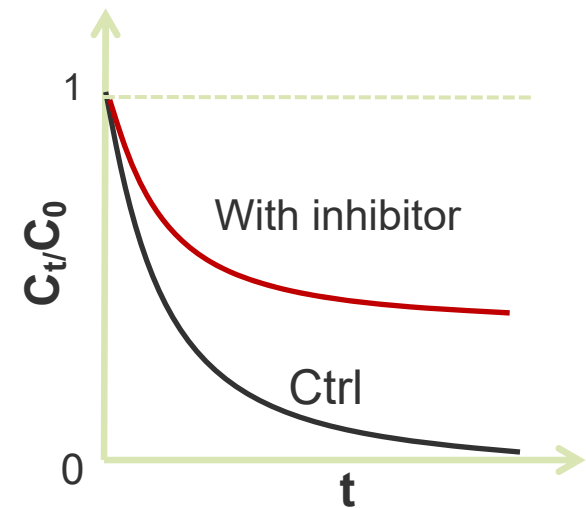
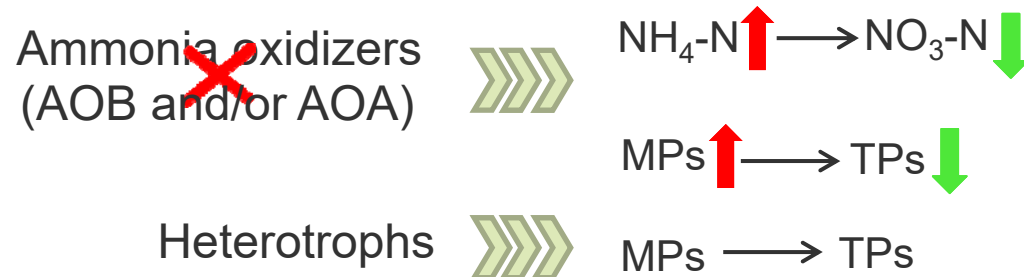
# Inhibition study

## Nitrifying Activated Sludge (NAS)

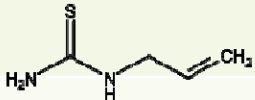
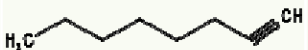
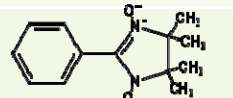


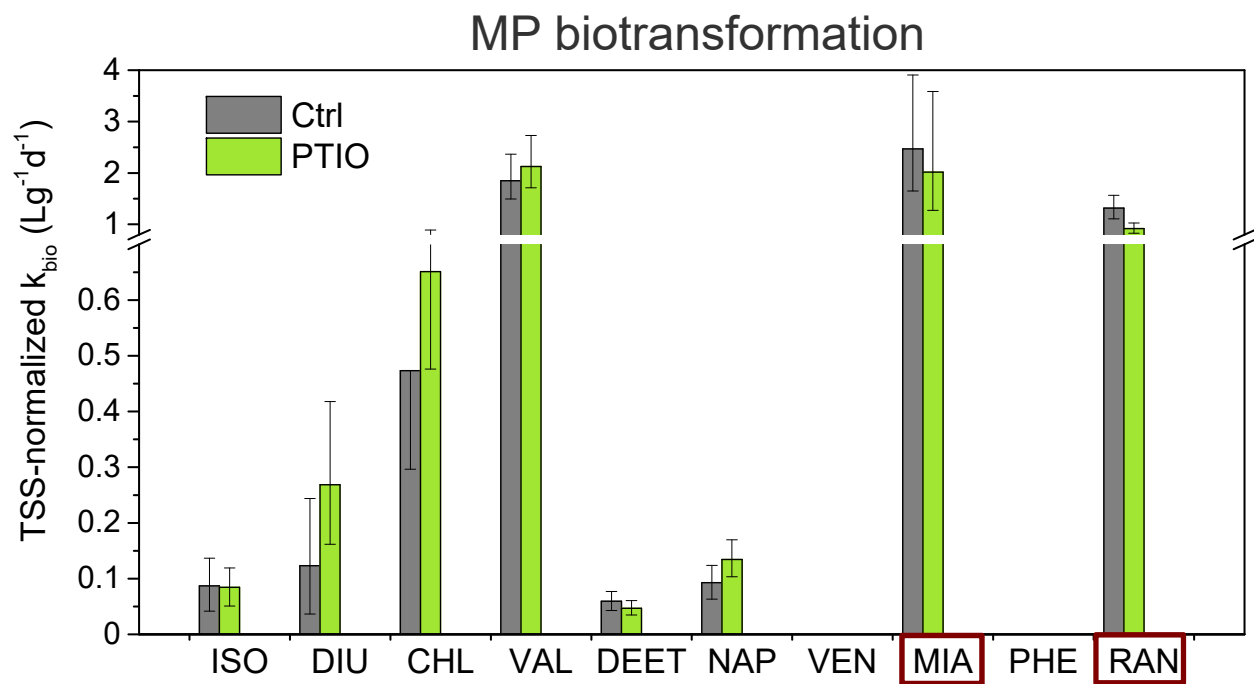
AUR: Ammونيا Uptake Rate

### *Hypothesis:*

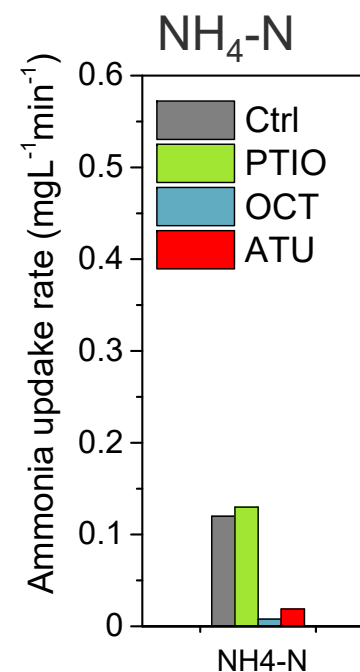


# Roles of AOA in MP biotransformation

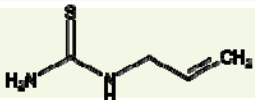
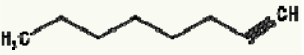
<i>Inhibitor</i>	<i>Structure</i>	<i>Conc.</i>	<i>Inhibition efficiency</i>	<i>Inhibition mechanism</i>	<i>Ref.</i>
Allylthiourea (ATU)		170 $\mu\text{M}$	AOB: 100% AOA: partial/none	Cu chelator	Ginestet <i>et al.</i> 1998, Jantti <i>et al.</i> 2013
Octyne (OCT)		20 $\mu\text{M}$ C <sub>aq</sub>	AOB: 100% AOA: partial/none	Covalent binding to AMO	Taylor <i>et al.</i> , 2013
PTIO		100 $\mu\text{M}$	AOB: none AOA: 100%	NO scavenger	Martens-Habbena <i>et al.</i> , 2015

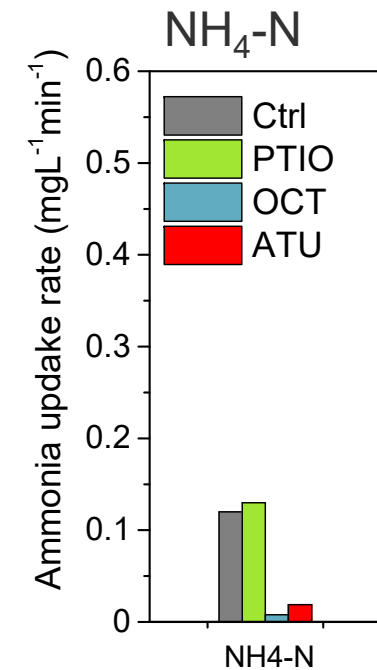
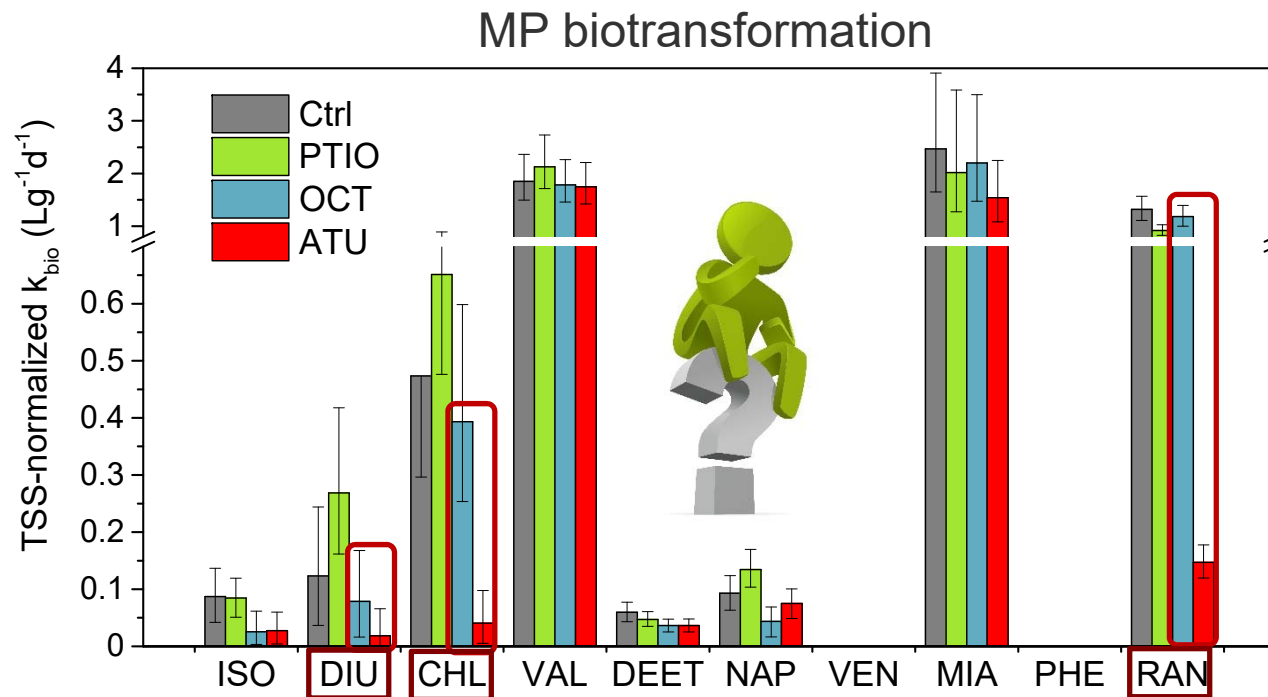


(Men *et al.*, 2016. ES&T)



## ATU and OCT exhibited different inhibition on AOB

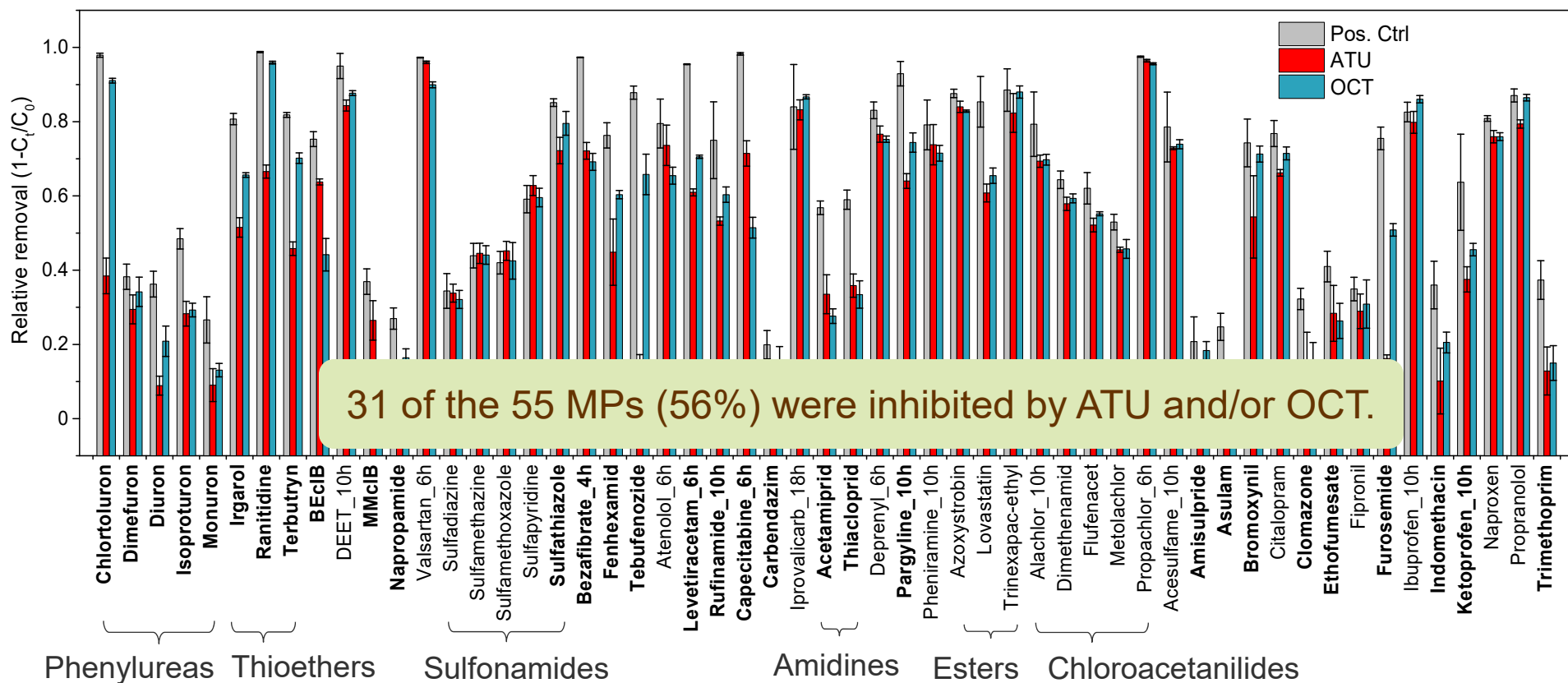
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# ATU exhibited higher inhibition for an extended MP list

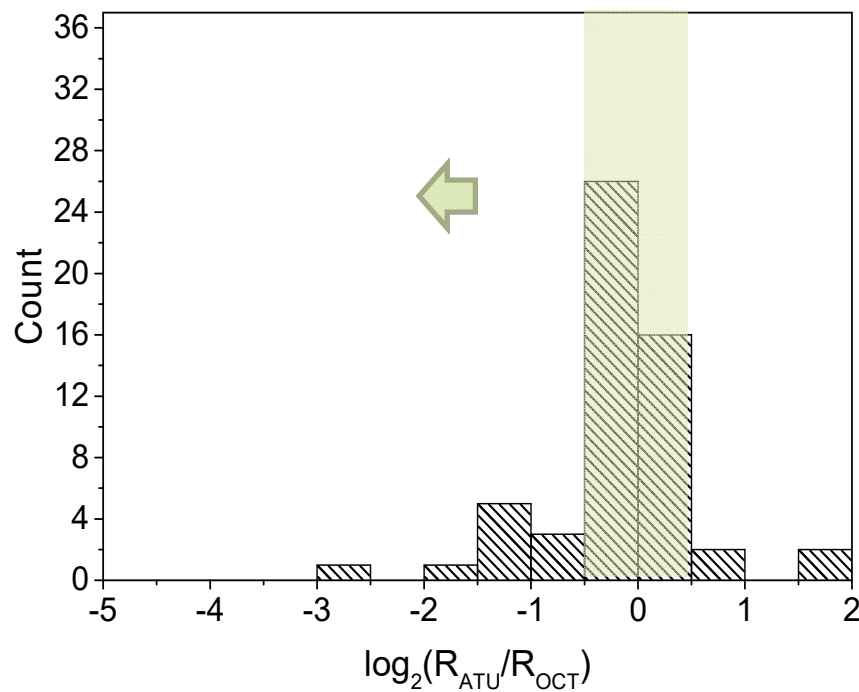
Relative Removal > 20%	Relative removal <20%	Strong sorption & fast abiotic removal	Total	AOB Inhibitors	Structure
55	20	4	79	Allylthiourea (ATU)	<chem>NC(=S)NCC=C</chem>
				Octyne (OCT)	<chem>CCCCCCCCC#C</chem>



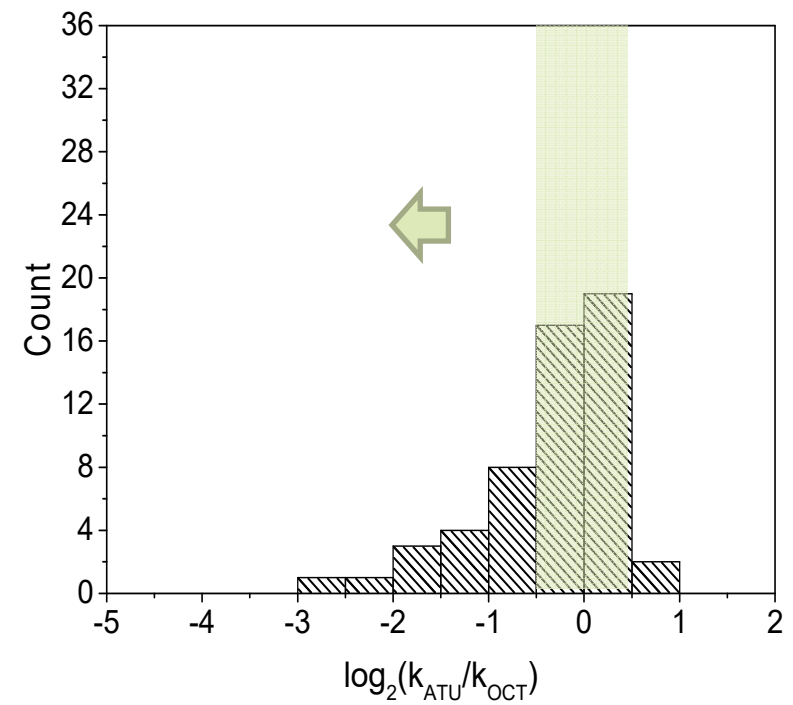
# 17 MPs with higher inhibition by ATU than OCT

ATU vs OCT

Removal (R)



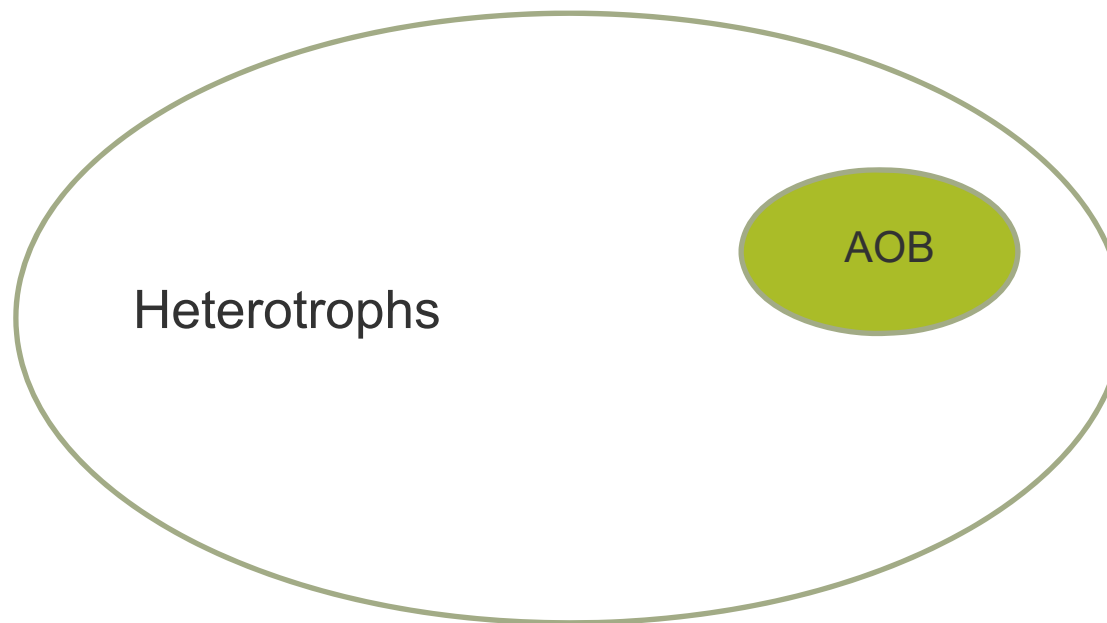
Rate constant (k), h<sup>-1</sup>



# Roles played by different NAS community members

Previous hypothesis: ATU is a specific AOB inhibitor.

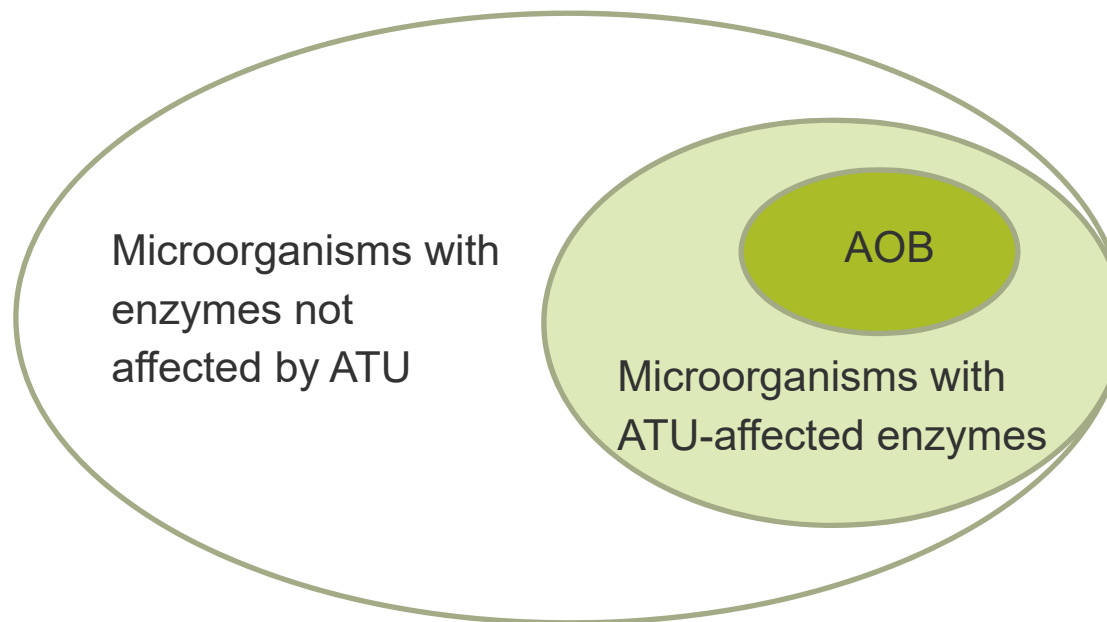
*Classification of members in a NAS community based on ATU sensitivity*



# Roles played by different NAS community members

This study: ATU does not specifically inhibit AOB, and other ATU-affected enzymes might also contribute to the biotransformation of some MPs.

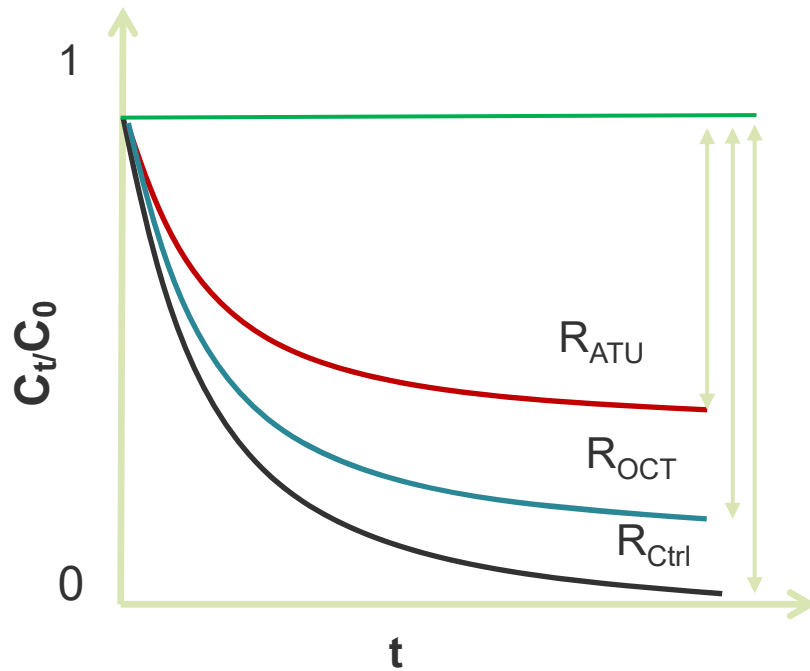
*Refined Classification of members in a NAS community based on ATU sensitivity*



Comparing inhibition effects of ATU and OCT

AOB	Other ATU-affected microbes	ATU-unaffected microbes
Similar inhibition	Higher inhibition by ATU	No inhibition

# Contribution of the three microbial groups



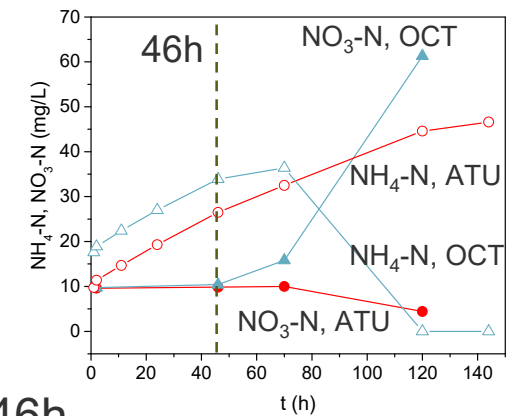
$$\text{Extent of inhibition } (I) = \frac{R_{Ctrl} - R_{Treated}}{R_{Ctrl}},$$

R: relative removal ( $1 - C_t/C_0$ )

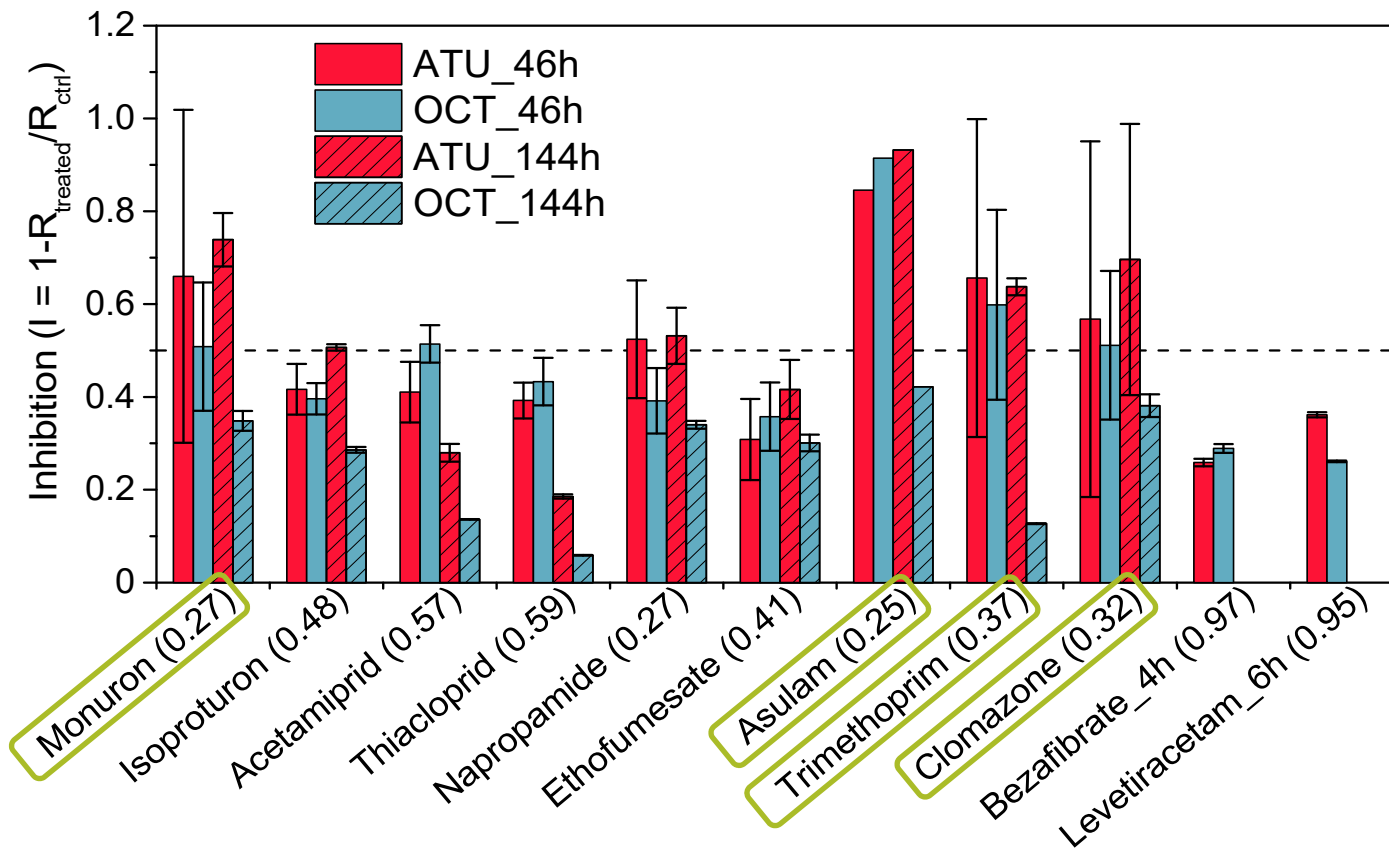
$I$  : how much the inhibited member contribute to MP biotransformation

# 11 MPs with similar inhibition by ATU and OCT

Recovery of nitrification after 46h in OCT-treated samples

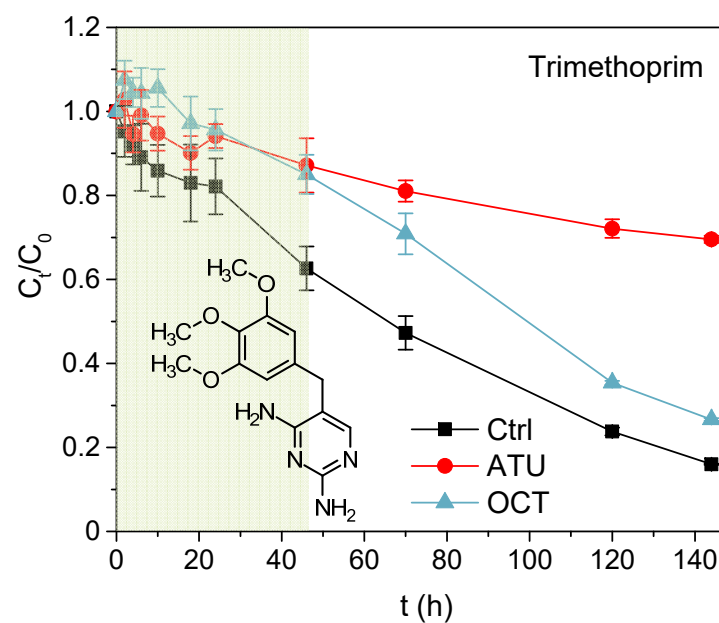
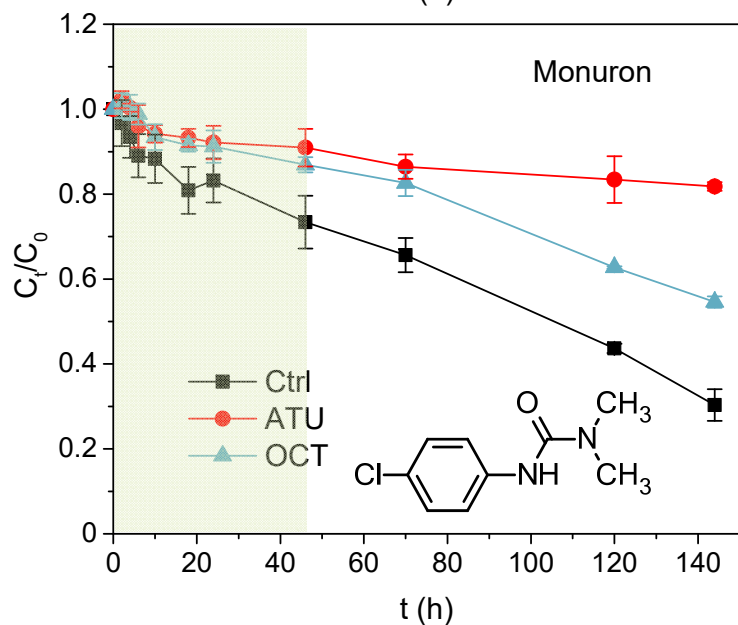
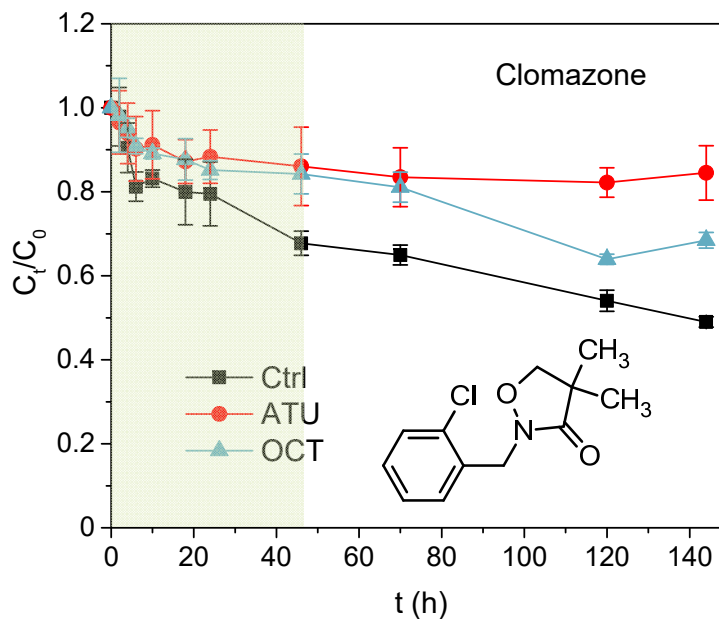
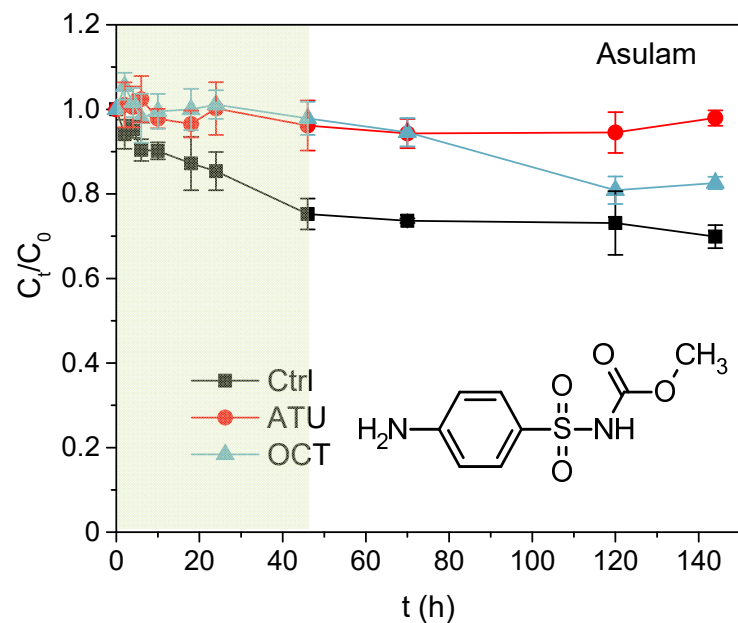


Recovery of biotransformation for the 11 Compounds after 46h



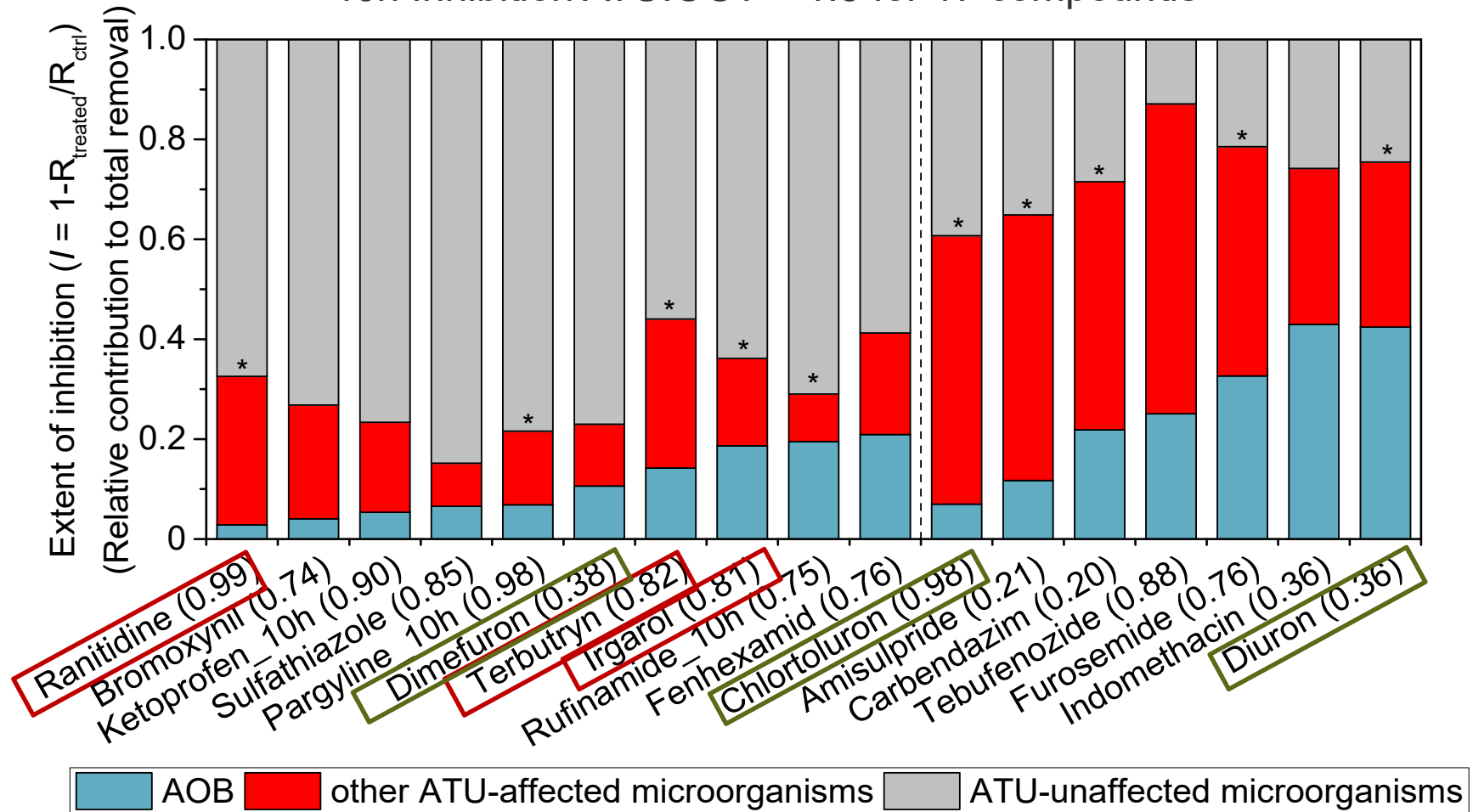


# AOB played a major role in the biotransformation of 4 MPs



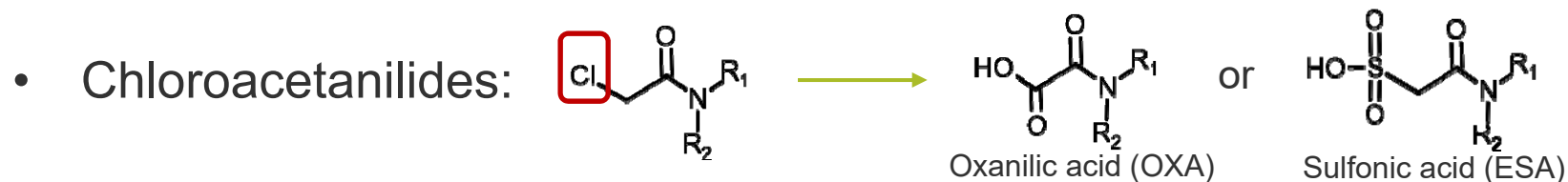
# 17 MPs with higher inhibition by ATU than OCT

46h Inhibition ATU:OCT > 1.5 for 17 compounds

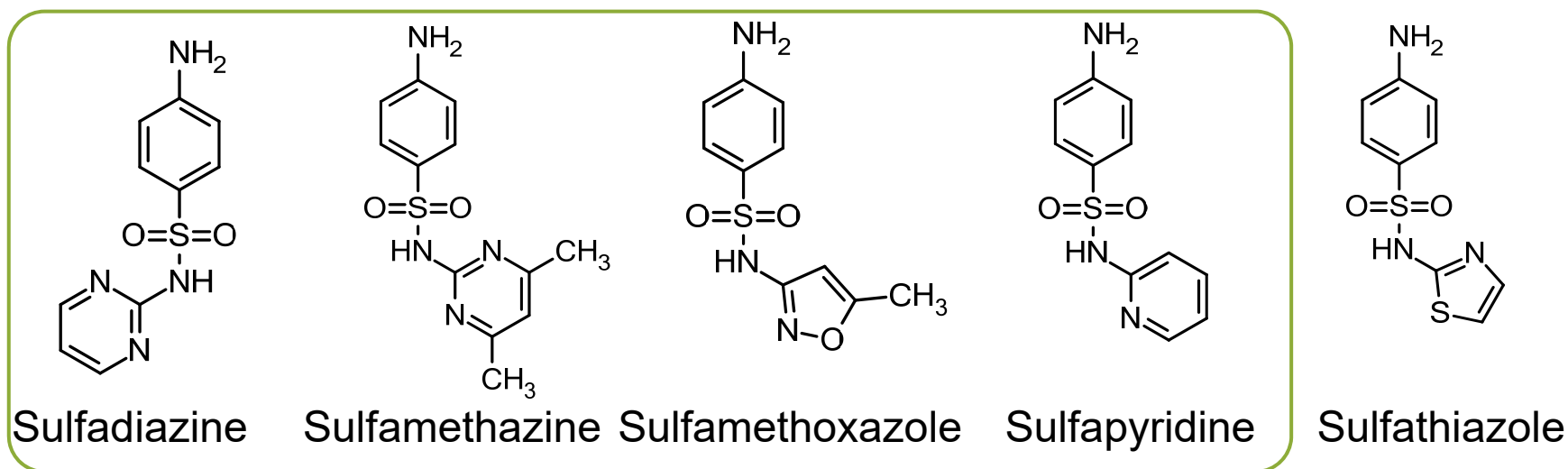


## No inhibition by the two AOB inhibitors for the following MPs:

- Esters: hydrolysis



- Sulfonamides: hydroxylation on aromatic ring



No inhibition

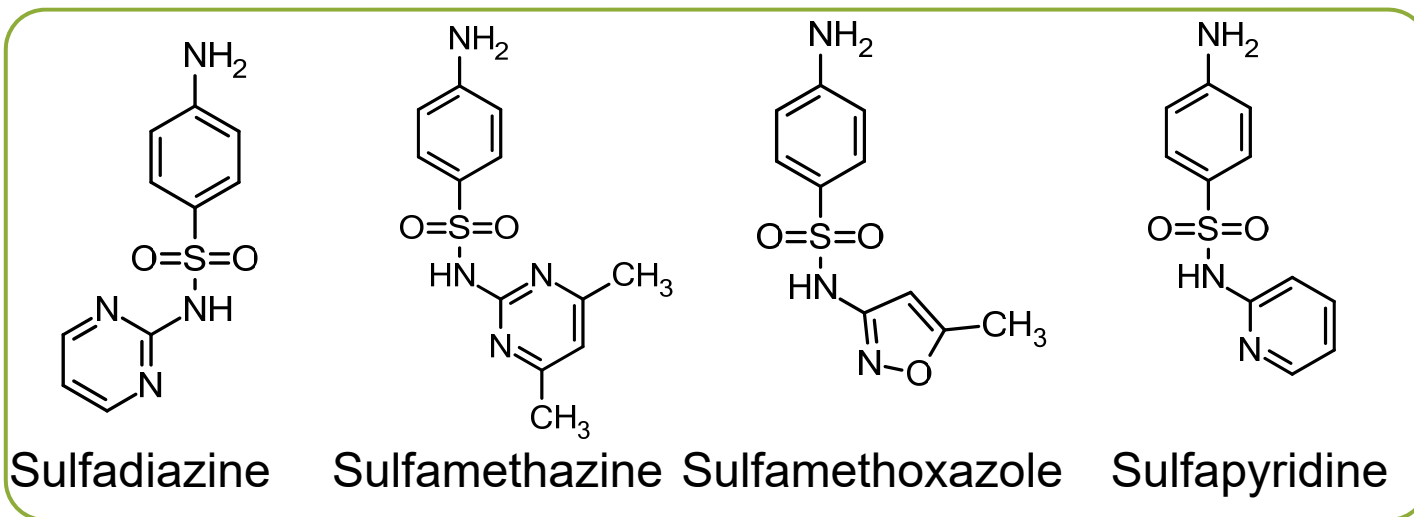
Inhibited by ATU  
(~ 15%)

# Compound specificity in AOB-related biotransformation

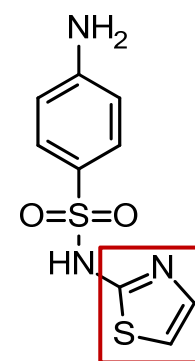
Phenylurea pesticides

Thioethers

## Sulfonamide antibiotics



No inhibition



Sulfathiazole

Inhibited by ATU  
(~ 15 %)

# Comparison with ATU inhibition studies in literature

MP	ATU added (mg/L)	TSS (g/L)	NH <sub>4</sub> -N added (mg/L)	MP conc. (µg/L)	<i>I</i> <sub>ATU</sub> in reference <sup>a</sup>	<i>I</i> <sub>ATU</sub> ( <i>I</i> <sub>oct</sub> ) in this study <sup>b</sup>	Reference
Ibuprofen	10	1	100	100	0.25	0.03 (0)	(Tran et al., 2009)
Ibuprofen	10	1 <sup>e</sup>	70/140	2000	0	0.03 (0)	(Roh et al., 2009)
Ibuprofen	12	1.8-5.1	<2/10/29	100	0-0.47	0.03 (0)	(Falas et al., 2012)
Atenolol <sup>c</sup>	30	1.73	20	15	0.63	0.04 (0.06)	(Sathyamoorthy et al., 2013)
Bezafibrate	50	4-16	20	1	0.48	0.26 (0.29)	(Maeng et al., 2013)
<b>Ketoprofen</b>	5	NA	0-0.5	2	0.76	0.41 (0.28)	(Rattier et al., 2014)
<b>Ketoprofen</b>	10	1	100	100	0.57	0.41 (0.28)	(Tran et al., 2009)
<b>Ketoprofen</b>	50	4-16	20	1	0.75	0.41 (0.28)	(Maeng et al., 2013)
<b>Diclofenac<sup>d</sup></b>	10	1	100	100	0.67	0.97 (0.58)	(Tran et al., 2009)
<b>Diclofenac</b>	50	4-16	20	1	0.32	0.97 (0.58)	(Maeng et al., 2013)
<b>Furosemide</b>	5	NA	0-0.5	2	0.78	0.79 (0.33)	(Rattier et al., 2014)
<b>Indomethacin</b>	10	1	100	100	0.86	0.74 (0.43)	(Tran et al., 2009)
<b>Irgarol</b>	11.1	2.5-4.7	50	1	0.37	0.36 (0.19)	(Margot et al., 2016)
<b>Terbutryn</b>	11.1	2.5-4.7	50	1	0.53	0.44 (0.14)	(Margot et al., 2016)
Trimethoprim	5	3.3	50	250	0.64	0.66 (0.60)	(Batt et al., 2006)
Trimethoprim	5	NA	0-0.5	2	<0	0.66 (0.60)	(Rattier et al., 2014)
Trimethoprim	10	NA	~24	0.5	0	0.66 (0.60)	(Khunjar et al., 2011)

# Insights into roles of AOB in MP biotransformation

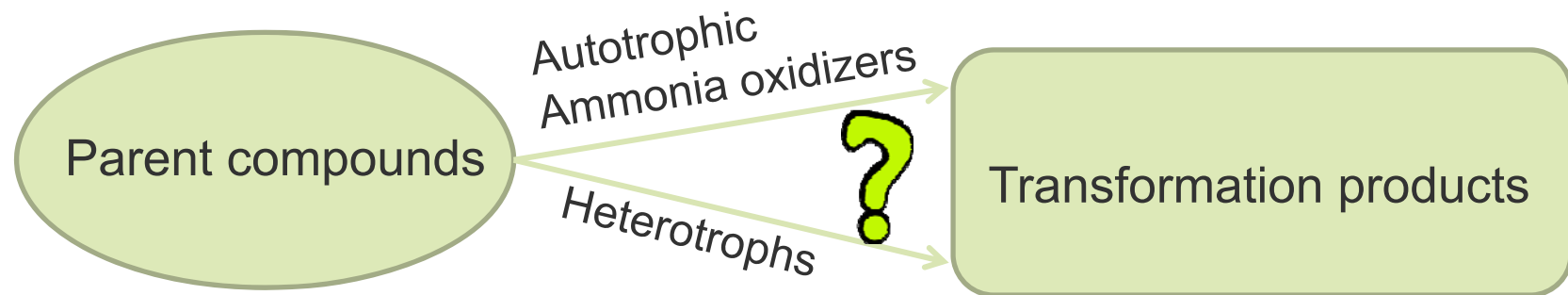
Microorganisms with enzymes not affected by ATU	Microorganisms with ATU-affected enzymes other than AOB	AOB
<i>This study</i>		
<ul style="list-style-type: none"> <li>• Esters</li> <li>• Chloroacetanilides</li> <li>• Sulfonamide antibiotics (except sulfathiazole)</li> <li>• Others</li> </ul>	<ul style="list-style-type: none"> <li>• Phenylureas</li> <li>• Thioethers</li> <li>• Others</li> </ul>	<ul style="list-style-type: none"> <li>• Asulam</li> <li>• Clomazone</li> <li>• Monuron</li> <li>• Trimethoprim</li> </ul>
<i>Literature</i>		
<ul style="list-style-type: none"> <li>• ✓ Ibuprofen</li> <li>• ✓ Naproxen</li> <li>• ✓ Valsartan</li> <li>• ✓ DEET</li> <li>• ✓ Trimethoprim</li> </ul>		<ul style="list-style-type: none"> <li>• ✓ Trimethoprim</li> <li>• Ketoprofen</li> <li>• Furosemide</li> <li>• Indomethacin</li> <li>• Irgarol</li> <li>• Terbutryn</li> </ul>





# Hypothesis

Causal relationship between ammonia oxidizers and MP biotransformation

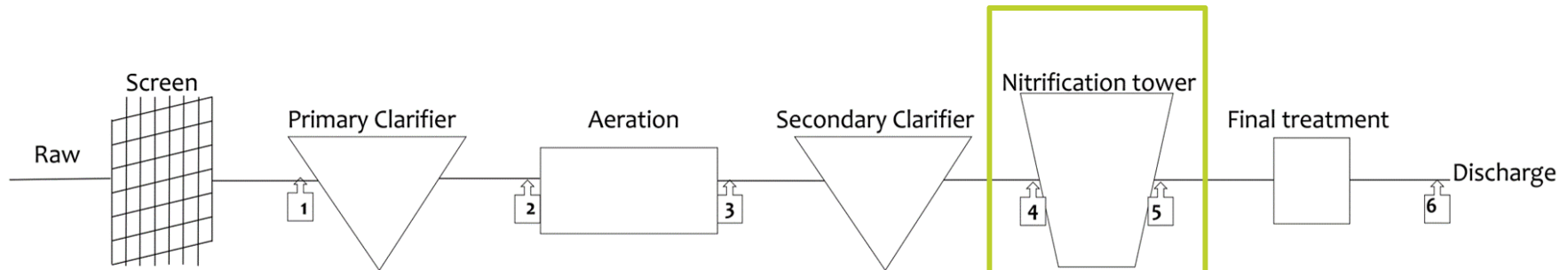


## To test the hypothesis

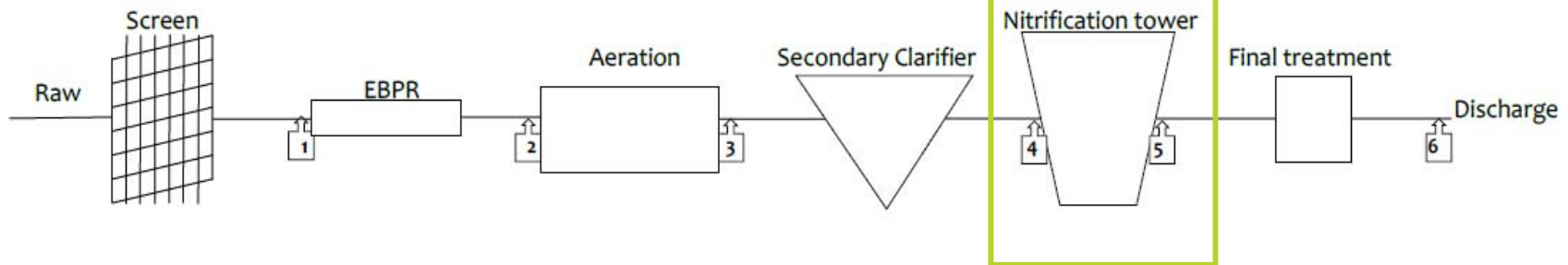
- ☐ Pure culture study
  - ✓ AOB
  - ✓ AOA
- ☐ Inhibition study
- ☐ Removal in full-scale WWTPs

# Roles of additional nitrification treatment (nitrification tower) in MP biotransformation

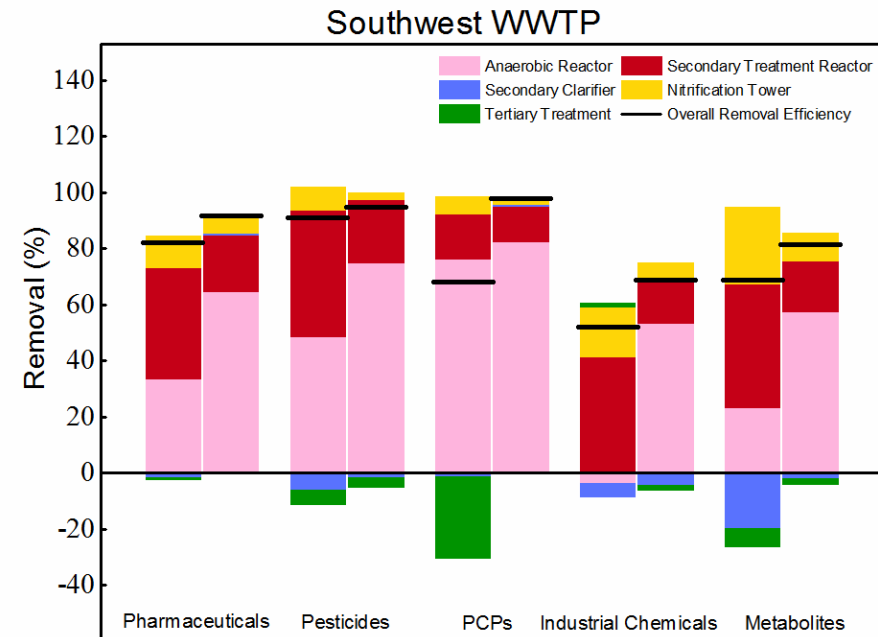
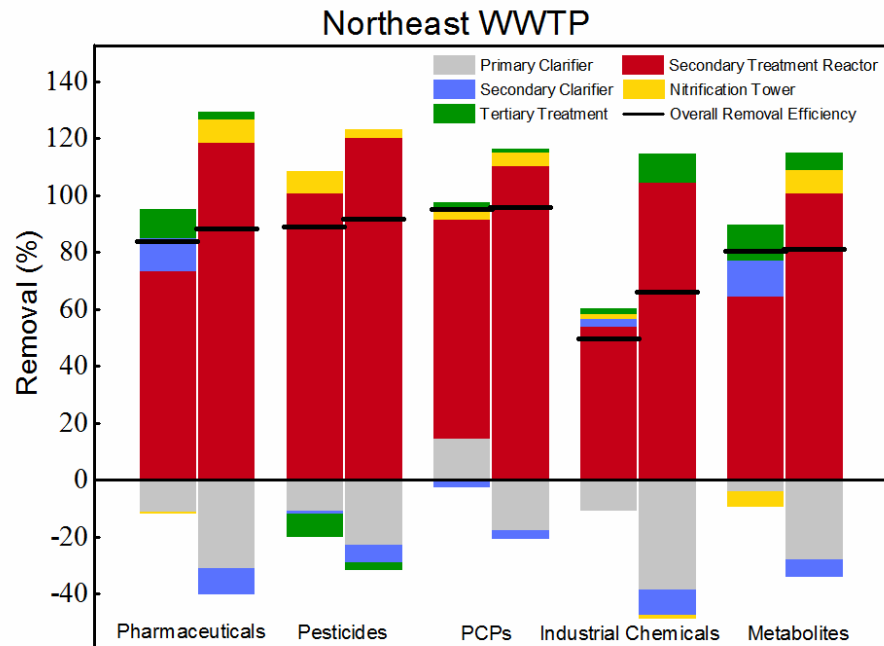
## Northeast WWTP



## Southwest WWTP



# MP removal in each treatment step



## Concluding Remarks

- ✓ AOB and AOA species were able to biotransform mianserin and ranitidine, likely via cometabolism by AMO.
- ✓ ATU is not a specific inhibitor to study the roles of ammonia oxidizers in NAS. Roles of AOB in MP biotransformation seem to be **overestimated** in inhibition studies using ATU. More specific inhibitors should be considered.
- ✓ AOB in NAS played a major role in the biotransformation of four compounds (i.e. **asulam, clomazone, monuron, and trimethoprim**).
- ✓ **Other microbes with ATU-affected enzymes** were strongly involved in the biotransformation of a number of MPs, including **thioethers** and **phenylureas**.
- ✓ AOB and microorganisms with ATU-affected enzymes were not involved in the biotransformation of **esters, chloroacetanilides** and **sulfonamide antibiotics (except for sulfathiazole)** investigated in this study.
- ✓ MP biotransformation was carried out by **multiple community members** in NAS. **TP identification** can further help differentiate roles of community members.





*Thank you for your attention!*

